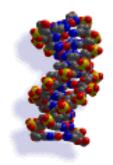




## DNA Analysis - Basic Tools and Techniques



## Zlatko Liber

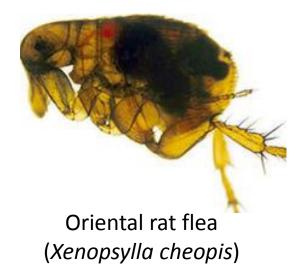
University of Zagreb, Faculty of Science, Zagreb, Croatia Centre of Excellence for Biodiversity and Molecular Plant Breeding, Zagreb, Croatia E-mail: zlatko.liber@biol.pmf.hr Why are archaeologists interested in studying ancient DNA (aDNA)?



King Richard III of England (1452–1485)

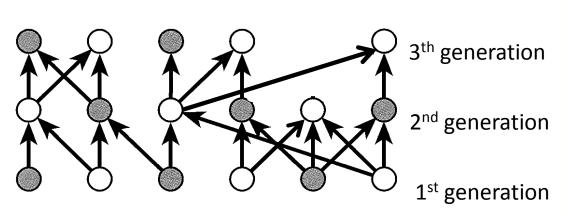


Vindija cave (Croatia)

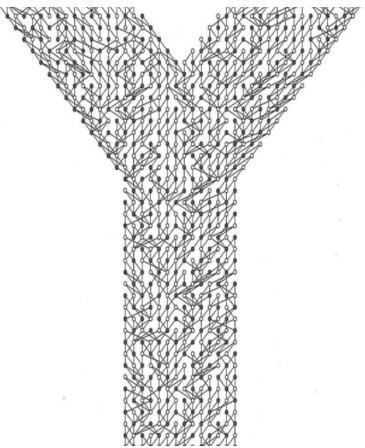


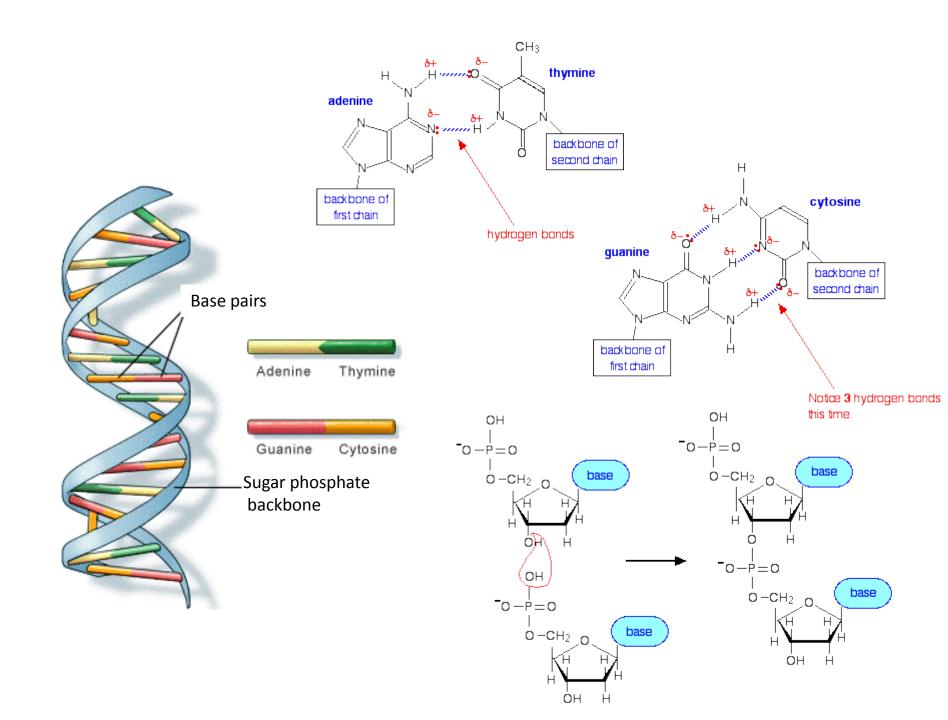
## Why are DNA molecules so informative?

**D**eoxyribo**N**ucleic **A**cid (**DNA**) is a molecule that carries all instructions used in the growth, development, functioning and reproduction in all known living beings.

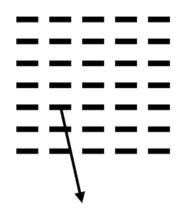


DNA is a time traveller!



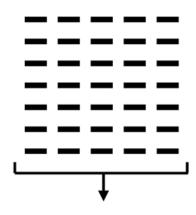


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one fragment (ca 1000 bp long) per run

2nd and 3rd generations of DNA sequencers (last ten years)



millions of fragments per run



(human genome project /1990 – 2003/)



MinION

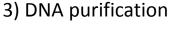
#### How to investigate DNA?

- 1. DNA isolation
- 2. PCR methods
- 3. Sequencing (1st, 2nd or 3th generation)
- 4. Bioinformatics

#### **DNA** isolation

#### 1) Cell lysis

2) Removing proteins and some other compounds3) DNA purification





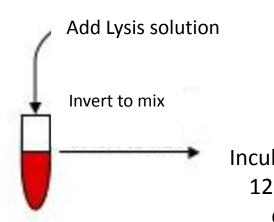






Archaeological sample need specific pretreatment because there is high possibility of contamination from microenvironment of the fossil, but also present day humans !!!

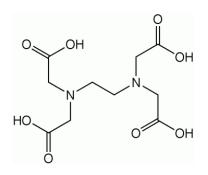




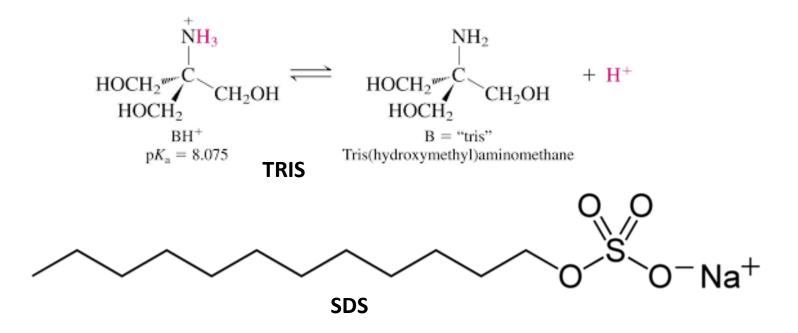


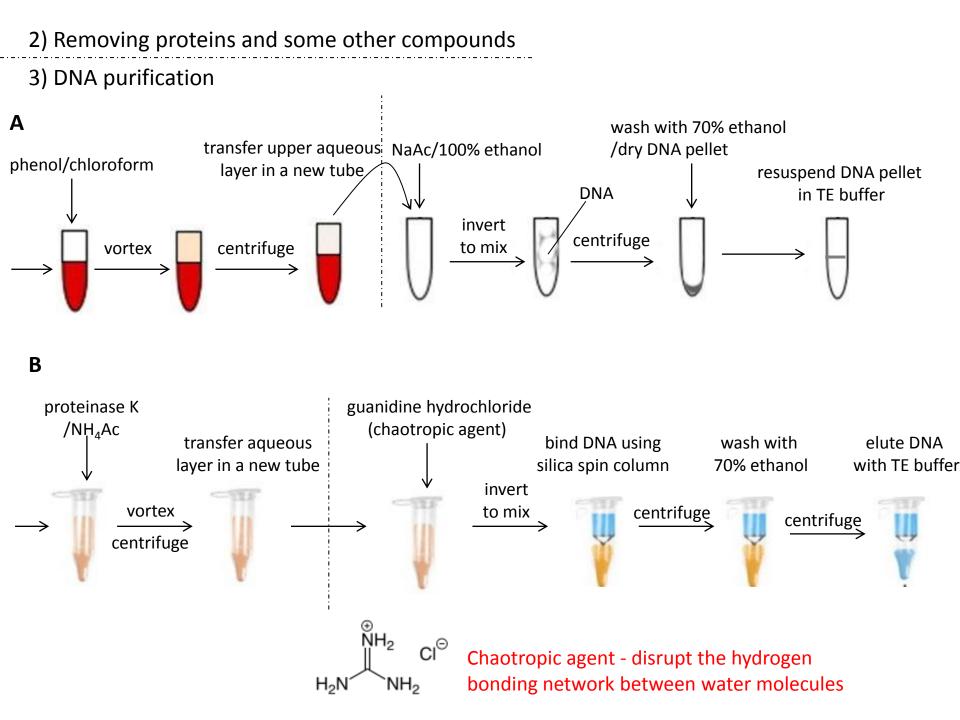
Incubate at 37 - 65 °C 5' to 12h with occasional or continuous stiring

EDTA



General solution of lysis buffer contains: TRIS-HCl, EDTA, SDS or CTAB





• DNA sample from two ~40,000-year-old Austrian cave bears was a mixture of bacterial, fungal, plant and other DNAs. Less than 6% of the recovered DNA was determined to be of cave bear (J. P. Noonan et al. 2005: Genomic sequencing of Pleistocene cave bears. Science 309, 597–599. )

The precautions usually applied in ancient DNA laboratory against DNA contamination:

- 1) complete separation of ancient laboratory and their rooms from other laboratories
- 2) direct delivery of all equipment and reagents to the laboratory
- 3) positive pressure generated with filtered air that excludes particles larger than 0.2 µm
- 4) UV irradiation and bleach treatment of all surfaces
- 5) bone surface was removed prior to extraction





#### **PCR** methods

#### Polymerase Chain Reaction

 amplifying a specific DNA fragment *in vitro* up to billion copies (Kary B. Mullis - Nobel Prize in Chemistry 1993)

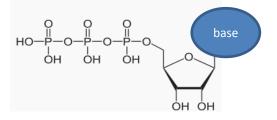


Thermal cycler device is able to specificaly increase and decrease temperature

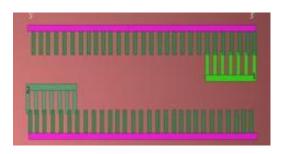
PCR ingredients:

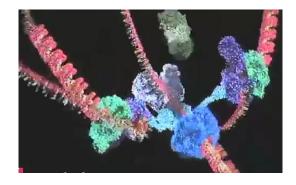
- 1) DNA
- 2) PCR buffer with MgCl<sub>2</sub>
- 3) Taq DNA polymerase
- 4) dNTP mix (A, C, G, T)
- 5) Primer 1
- 6) Primer 2

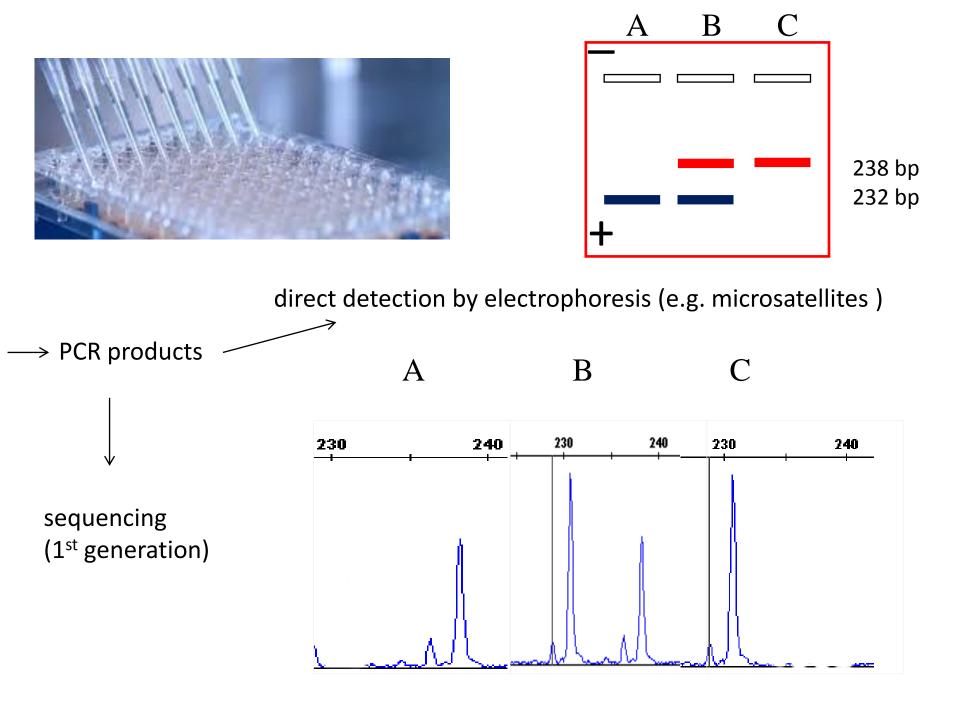


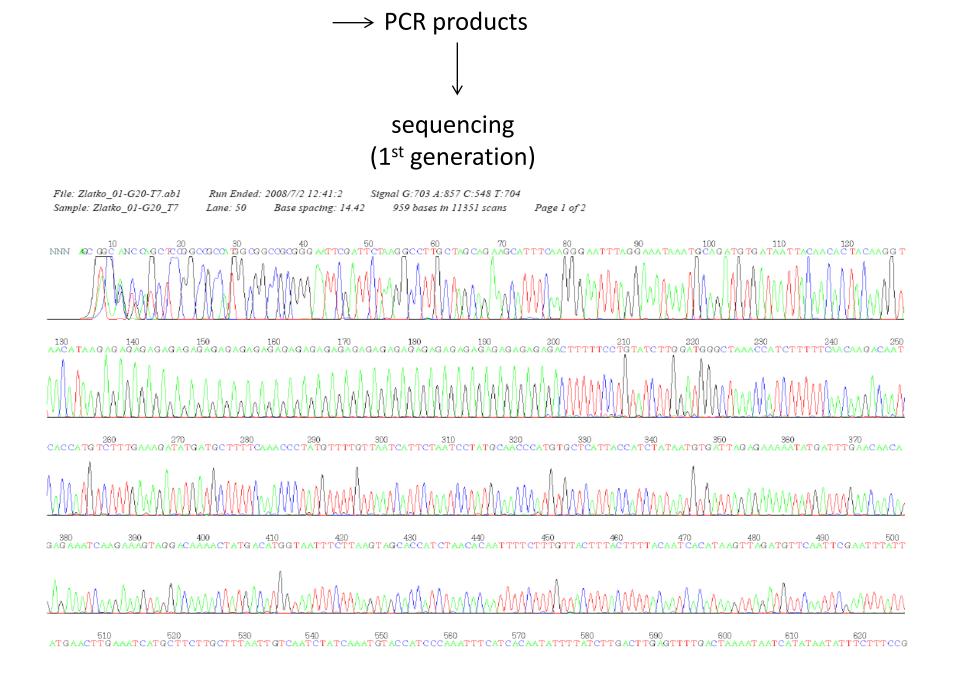






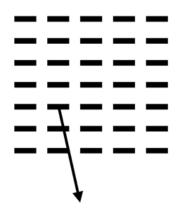






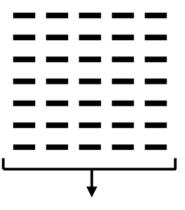
#### **NGS** sequencing

1<sup>st</sup> generation of DNA sequencers (started 1987)



one fragment (ca 1000 bp long) per run

2<sup>nd</sup> and 3<sup>rd</sup> generations of DNA sequencers (last ten years)

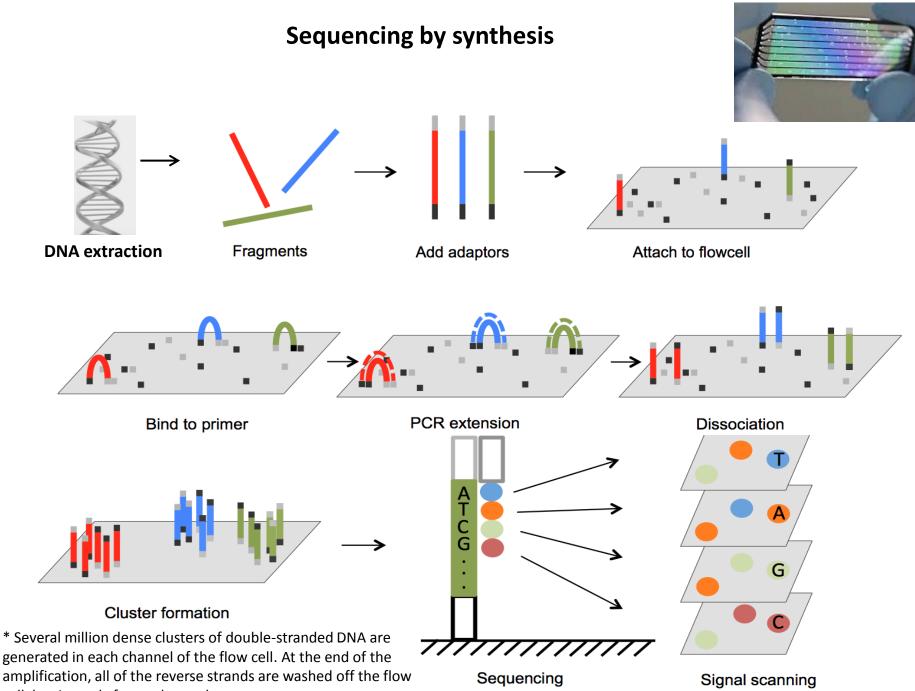


millions of fragments per run

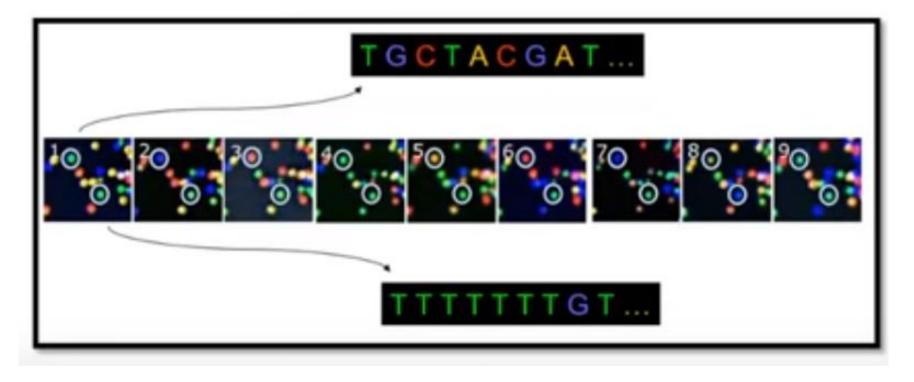
#### NGS = Next Generation Sequencing

1) Sequencig by synthesis

2) Nanopore sequencing



cell, leaving only forward strands.





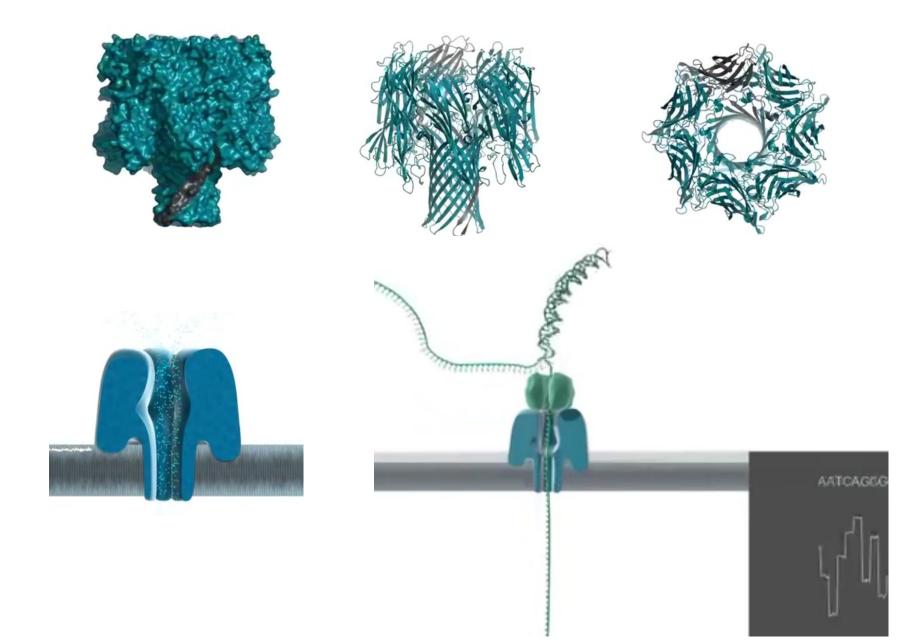
Illumina NGS sequencers (sequencing by synthesis)

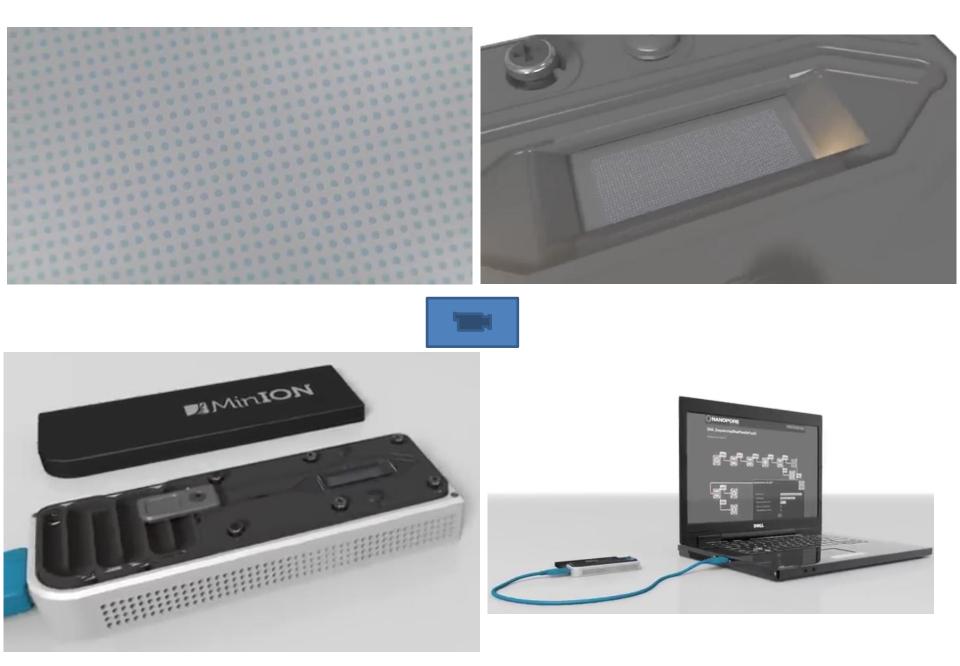
# **MiSeq Series** MAX OUTPUT 15 Gb MAX READ NUMBER 25 million MAX READ LENGTH

2×300 bp

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CCTGGGAAGGGTCAGTGCGAGTGAGGGCCCTGG AGGGTCAGTGC	TG	GTGAGGGCCCTGGGAAGGGTCA GAGTGAGGGCCCTG
cccgggacgggtcagtgcgggtgagggccctgggaagggtcagtgc	tg	dtgagggccctgggaagggtcagtgcg gagggccctg
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GGAAGGGTCAGTGCGAGTGAGGGCCCTGGGAAGGGT	10	GAGGGCCC GGGAAGGG CAG GCGCG GAGT TG
CCTGGGAAGGGTCAGTGCGAGTGAGGGCCCTGGGAAGGGTC	tg	gt gagggccc t gggaagggt cagt gcgag t gagggccc t g
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CCTGGGAAGGTTCAGTGCGAGTGAG CCCTGGGAAGGGTCAGTGC	16	GAGGGCCC GGGAAGGG CAG GCGAG GAGGGCCC G
	16	

#### Nanopore Sequencing





• yield : 21 Gb, read number 2.2 millions, longest read: 200Kb, run time 48h,, weight 87g